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Review Article

Caffeic acid phenethyl ester (CAPE): pharmacodynamics and potential for therapeutic application

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Abstract

Caffeic acid phenethyl ester (CAPE) is the major pharmacologically-active component of some propolis types, rich in polyphenols, such as poplar propolis types. CAPE has the potential to be applied as a pharmaceutical as it possesses most of the pharmacological activities of propolis, such as anti-proliferative, antioxidant, immunomodulatory, antidiabetic, anti-inflammatory and antimicrobial. Its advantage is that it lacks some of the downsides of total propolis extracts, such as inability for unified standardization, which is cornerstone for implementing its therapeutic potential as a drug. The current paper provides an overview on the pharmacodynamic principles of CAPE. We present literature search outcomes form ClinicalTrials.gov database and from scientific publications, available on Scopus and Crossref databases. We take a round view of CAPE's potential therapeutic implications in light of approved drugs with related modes of action.

Keywords

Pharmacology, therapeutic application, selective inhibitor of nuclear export, NF-κB activation inhibition

Introduction

CAPE is a major active constituent of some types of propolis, rich in polyphenolics (Murtaza et al. 2014, Zhang et al. 2014, Tolba et al. 2016, Bankova et al. 2018). CAPE is the most studied individual component of propolis at present (Bankova et al. 2018). It has been shown to possess most of the reported biological activities, characteristic to total propolis extracts (Zabaiou et al. 2017), including antimicrobial (Arasoglu et al. 2016), antiproliferative (Akyol et al. 2013), protective against oxidative stress-mediated tissue damage, anti-inflammatory, immunomodulatory (Turan et al. 2015) and antidiabetic (Celik et al. 2009).

Pharmacological mechanisms

Studies in the first decade after the discovery of CAPE proved fruitful in confirming that it possesses broad biological activities, mostly coinciding with those of whole propolis extracts. The cornerstone studies during this stage of CAPE research were introduced in the first part of the current article. CAPE was shown to be most effective as an antineoplastic, antioxidant and anti-inflammatory agent and as inhibitor of some enzymes. This knowledge provided the basis for a boost of research articles in CAPE's underlying mechanisms of action. CAPE was tested on diverse *in vitro* and *in vivo* disease models. Data

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on its biological activities has been systematized in extensive review articles regarding its antineoplastic (Ozturk et al. 2012, Akyol et al. 2013, Lin et al. 2013, Kuo et al. 2015, Watanabe and Sforcin 2018), antioxidant and protective towards drug-induced adverse reactions (Akyol et al. 2014, Magnani et al. 2014, Erdemli et al. 2015, Tolba et al. 2016, Anjaly and Tiku 2018), anti-inflammatory (Tolba et al. 2013, Armutcu et al. 2015, Murtaza et al. 2014), antidiabetic (Pittalà et al. 2018) and antiviral (Erdemli et al. 2015) activities. Here, we are going to highlight some therapeutically-relevant pharmacological mechanisms of CAPE. For convenience, we are about to group them as mostly relevant to signal transduction modulation, direct enzyme inhibition and direct antioxidant activities, although CAPE's activity is most probably a combination of the mentioned mechanisms.

Signal transduction modulation

The source of evidence for the influence of CAPE on signal transduction pathways is predominantly research on its cytotoxic effects on different tumor cell lines. The reason behind the selective cytotoxicity of CAPE towards cells that have undergone chemical/viral transformation compared to non-transformed cells (Grunberger et al. 1988) has been related to its inhibition of the activity of nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) (Natarajan et al. 1996). NF- κ B is a transcription factor, with a central role in the regulation of different cell functions as cell survival, inflammation, stress response and metabolism as well as tissue development homeostasis and differentiation (Karin 2009, 2012, Aggarwal and Sung 2011, Baker et al. 2011, Lingappan 2018). In the inactive state, NF-KB proteins are retained in the cytoplasm by association with inhibitor of kB (IkB) proteins. NF-kB is activated when IkB becomes phosphorylated by IkB kinases (IKKs), which ultimately lead to its degradation by the proteasome. This in turn leads to the release of the uninhibited, active NF-KB proteins, which can then translocate to the nucleus and exert their effect by activating the transcription of proteins (Moynagh 2005). The state of activation of NF-KB is paramount for the regulation of many signal pathways, which could lead either to cell survival or call death, by activation of caspases and apoptosis or other cell death mechanisms. The interaction between signaling pathways and the fate of the cell depend on the intra- and extracellular biochemical context. An example of such signaling is tumor necrosis factor (TNF), which is one of the strongest inducers of NF-kB activity. TNF functions as a pro-apoptotic factor and an inflammatory cytokine and depending on the biochemical context can either cause apoptosis or cell survival and proliferation. It is generally apoptotic only when NF-KB is blocked (Flusberg and Sorger 2015). The exact mechanism of CAPE's interaction with components of NF-KB-mediated signal transduction is not ultimately clarified. However, CAPE has been shown to block NF-κB activation in tumor (Natarajan et al. 1996, Watabe et al. 2004, Liu et al. 2018) and immune cells (Lalor

et al. 2007, Wang et al. 2010). It has been shown to prevent the p65 (one of the active components of NF- κ B) from translocation in the nucleus and from binding to DNA (Natarajan et al. 1996, Liu et al. 2018). It has been speculated that CAPE directly inhibits IKK (Lee et al. 2010). Another study infers that the cellular target of CAPE is Exportin 1/ chromosomal maintenance 1 (XPO1/CRM1), a nuclear transporter protein which functions as a nucleocytoplasmic exporter of growth-regulatory factors. CAPE targets specifically a non-catalytic cysteine 528 of XPO1/ CRM1, which results in inhibition of its function. (Wang and Liu 2019). Among the target proteins of XPO1/CRM1 are NF-κB, Wnt/β-catenin, PI3K/Akt, p53, FOXOs (Forkhead box proteins), etc. (Wang and Liu 2019). It has been reported that XPO1/CRM1 inhibitors are among the anticancer agents with the broadest spectrum of activity towards diverse malignancies (Wang and Liu 2019). They exert anti-inflammatory effects in preclinical models of neurodegeneration (Haines et al. 2015), which could be also the mechanism of the antineoplastic and anti-inflammatory effects of CAPE. Whether the primary pharmacological mechanism of CAPE is inhibition of XPO1/CRM1 or of NF-kB activity (by inhibition of IKK, p65 translocation or DNA binding), there is non-clinical data, showing that CAPE affects signaling pathways, that are known to be in interplay with all of the discussed mechanisms. For example, CAPE enhances the expression of the tumor suppressor protein p53 in glioma cells (Lee et al. 2003). The tumor suppressor p53 is a transcription factor, regulating DNA-repair and apoptosis. It is the most frequently mutated gene in human cancer (Khoo et al. 2014) and its crosstalk with NF-KB has been implicated in the pathogenesis of stress- and inflammation induced cancer (Schneider and Krämer 2011). CAPE also interferes with FOXO signaling by increasing the levels of the FOXO-1 downstream tumor suppressor in prostate cancer cells (Tolba et al. 2013). FOXO proteins are important transcriptional effectors to the insulin and IGF-1 signaling pathway, they promote the antioxidant defense of cells (Martins et al. 2016) and block NF-KB activity and inflammation (Salminen et al. 2008). In another study, CAPE suppressed TGF-beta induced Akt phosphorylation and the migration of adenocarcinoma cells (Shigeoka et al. 2004). It also inhibited Akt phosphorylation and NF-KB activation in CD4+ T cells (Wang et al. 2010). The PI3K-Akt pathway promotes survival. Its activation leads to NF-KB activation and has been reported to have a role in type 2 diabetes (Carracedo and Pandolfi 2008). Another study shows that CAPE suppressed canonical Wnt signaling of prostate cancer cells, reducing their invasiveness (Tseng et al. 2016). Wnt signaling pathway activation is related to carcinogenesis and promotion of metastasis (Zhan et al. 2017). It's dysregulation predisposes for metabolic dysfunction and type 2 diabetes (Fuster et al. 2015) and is known to interact with NF-KB signaling during inflammation (Ma and Hottiger 2016). Another signaling pathway, regulated by NF-KB and influenced by CAPE, is nuclear factor erythroid 2-related factor 2 (Nrf-2) (Wardyn et al. 2015, Kucukgul 2016). Nrf2 is a transcription factor with cytoplasmic location, which upon activation initiates the expression of genes, coding anti-oxidant, anti-inflammatory and detoxifying proteins, such as heme oxygenase 1 (HO1), superoxide dismutase (SOD), catalase (CAT) and glutathione-s-transferase (GST) (Keum 2012, Loboda et al. 2016). CAPE exerts its antioxidant effects through increased HO1 expression, mediated by Nrf-2 (Stähli et al. 2019). A probable mechanism of Nrf-2 activation by CAPE has been proposed. CAPE's catechol moiety is responsible for it binding to Kelch-like ECH associated protein 1 (Keap1), which keeps Nrf-2 inactive by anchoring it in the cytoplasm. It is supposed that the binding of CAPE to Keap1 results in translocation of Nrf-2 to the nucleus and its activation (Kim et al. 2013). The given examples show the importance of XPO1/CRM1 and NF-κB inhibition in the plethora of effects of CAPE on tumor cells, inflammation and supposedly in metabolic diseases as type 2 diabetes. Based on the presented research, CAPE could be classified as a selective inhibitor of nuclear export (SINE) or, informally as such a pharmacological group has not yet been defined, as an inhibitor of NF-KB signaling. Noteworthy, it has been shown that the mechanism of action of anticancer and anti-inflammatory drugs is related to NF-KB inhibition (Urushibara et al. 2004, Baud and Karin 2009, Gan et al. 2016). Although Baud and Karin (2009) hypothesize that a logical side effect of effective IKK/NF-kB inhibitors should be immunosuppression, a major pitfall could be the pronounced ability of such inhibitors to stimulate the production of interleukin 1 beta (IL-1 β) due to overactivation of the inflammasome during bacterial infections. Recently, the first in class SINE - Selexinor was granted accelerated approval by the US Food and Drug Administration (FDA) for "adult patients with relapsed or refractory multiple myeloma (RRMM) who have received at least four prior therapies and whose disease is refractory to at least two proteasome inhibitors, at least two immunomodulatory agents, and an anti-CD38 monoclonal antibody." (Center for Drug Evaluation and Research 2019). Its most common adverse reaction is thrombocytopenia, which is not related to direct cytotoxicity, but rather to delayed maturation of megacaryocytes (Machlus et al. 2017). The existence of approved drugs, with relevant mechanisms of action underlines the good prospects for the clinical application of CAPE in neoplastic and immunopathological conditions.

Target enzyme inhibition

Another group of pharmacodynamic mechanisms of potential clinical importance stem from the ability of CAPE to inhibit the activity of some enzymes. For example, CAPE has been shown to selectively inhibit human matrix metalloproteinase-9 (MMP-9) and matrix metalloproteinase-2 (MMP-2) with IC₅₀ values respectively 2 and 5 μ M, but it did not inhibit MMP-1, MMP-3, MMP-7 and cathepsin K (Cat K) (Chung et al. 2004). Another study reports that CAPE inhibits MMP-9 at even lower concentrations with IC₅₀ of 1–2 nM (Jin et al. 2005). The MMP family's functions are to degrade extracellular matrix – forming proteins. Abnormal MMP activity (of MMP-2 and MMP-9 particularly) has been detected in pathologies such as cancer metastasizing (Iochmann et al. 2009, Li et al. 2017). So far, due to musculoskeletal toxicity, related to lack of specificity (Skiles et al. 2004), only one MMP inhibitor drug has been approved – a subantimicrobial dose doxycycline (Periostat) (Caton and Ryan 2011). The inhibiting activity of CAPE towards MMP-2 and MMP-9 could be promising in dental practice (Table 1), but could also be effective in alleviating tissue damage due to cerebral ischemia and stroke (Dong et al. 2009).

Another enzyme that has been researched as a target for CAPE inhibition is human immunodeficiency virus type 1 (HIV-1) integrase (Erdemli et al. 2015). The lentivirus HIV essentially needs the integrase enzyme in order to transfer its genetic material into the host cell's DNA. HIV-1 integrase catalyzes two biochemically and temporally separate steps: the first step consists of cleavage of a 3' terminal dinucleotide of the virus DNA. The second step, catalyzed by HIV-1 integrase is strand transfer, resulting in the integration of virus DNA in specific loci in host DNA. (Andrake and Skalka 2015). It has been shown that CAPE selectively inhibits the second, integration step (Fesen et al. 1994). All approved drugs (first generation: Raltegravir and Elvitegravir; second generation: Dolutegravir), inhibiting the HIV-1 integrase also target the second reaction, that's why they are called integrase strand transfer inhibitors (InSTIs) (Dow and Bartlett 2014). InSTIs are effective when included in antiretroviral polypragmasy and as pre-exposure prophylaxis for individuals in high risk of infection (Günthard et al. 2016). The antiviral activity of CAPE, however is multimodal, as it influences other stages in the retroviral life cycle due to its effects on the signal transduction pathways in the host cell (Erdemli et al. 2015).

CAPE has been shown to inhibit the in vitro activity of the cyclooxygenases COX-1 and COX-2, enzymes that are a popular target for non-steroidal anti-inflammatory drugs. The IC₅₀ values were 58 μ M for both enzymes. However, in the in vivo situation, the effect of CAPE on COX activities may be enhanced by its ability to inhibit their gene expression (Michaluart et al. 1999). CAPE has also been shown to inhibit arachidonate 5-lipoxygenase (5-LOX)-catalyzed oxygenation of linoleic acid and arachidonic acid and it has been shown that it inhibits the enzyme by a complete uncompetitive mechanism, related to its antioxidant capacity (Sud'ina et al. 1993). 5-LOX is also an enzyme, which functions in the production of proinflammatory molecules (leukotrienes) and its inhibition is an effective strategy in the treatment of inflammatory disorders. An approved drug for the treatment of asthma, which is an inhibitor of 5-LOX is Zileuton (Aparoy et al. 2012).

CAPE also inhibits xanthine oxidase (XO) by means of competitive inhibition at low concentrations with an IC_{50} of 6.26 μ M (Wang et al. 2009). XO's metabolic function is related to purine degradation, but it can also generate reactive oxygen species (Berry and Hare 2004). A clinical trial with the XO inhibitor Allopurinol, used for the treatment

of high blood uric acid levels concludes that it acutely improves the levels of high-energy phosphates as adenosine triphosphate's (ATP) flux through creatine kinase (CK) in the heart of patients with heart failure, which could protect them from disease aggravation (Hirsch et al. 2012).

In order to better understand CAPE's pharmacological action and effectively advance it to the bedside, it is relevant to clarify whether stereoisomerism influences its effects on enzymes and proteins, as such information is unavailable in scientific research databases.

Direct antioxidant activity

Research on the antioxidant activity of CAPE shows that it is capable of neutralizing oxidative stress (Tolba et al. 2016). Oxidative stress is known to be an important etiological factor in age-related diseases, such as cardiovascular and neurodegenerative disease, also in diseases with an inflammatory component as chronic obstructive pulmonary disease, metabolic diseases and cancer (Liguori et al. 2018). It has been shown that CAPE is the major component of propolis, responsible for its antioxidant activities (Rossi et al. 2002). Presumably, its antioxidant properties are related to its aforementioned effects on signal transduction, immunosuppressive activity and inhibition of enzymes, related to oxidative stress. The same is valid for the activity of CAPE as a protective agent against drug-induced toxicity, mediated by oxidative stress-related toxic mechanisms (Murtaza et al. 2014). Moreover, it has been shown that small molecules with direct antioxidant capacity, such as vitamins and dietary polyphenols can potentially protect against oxidative stress-related pathology (Liguori et al. 2018). CAPE exerts strong antioxidant activity, which is evident from it being an effective radical scavenger in the DPPH (Choi et al. 2019), and ABTS antioxidant tests (Bak et al. 2016). The presence of catechol moiety, the double bond in CAPE's molecule and the high lipophylicity are structural factors, responsible for its excellent antioxidant activity (Razzaghi-Asl et al. 2013).

Clinical trials

CAPE has a simple chemical structure (Fig. 1) and possesses the most utilized pharmacological activities of propolis, some of which at very low concentrations (antiproliferative and antioxidant), therefore it could be utilized as a medicine with greater success than propolis total extracts. Although the aim of the current review is not to extensively evaluate clinical data on propolis, in order to exemplify the interest towards its pharmaceutical application and the abundance of such data, we did a quick search in one of the biggest clinical trials' databases – https://clinicaltrials.gov with the keyword "propolis" in the search feature box, where drug information should be included. The search (as of 1.7.2019) resulted in 40 hits, dominated by dental products, related to its antibacterial, anti-inflammatory and analgesic properties (21 hits), fol-

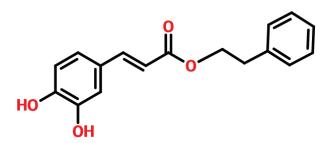


Figure 1. Chemical structure of CAPE.

lowed by diabetic foot treatments (4 hits), skin and wound conditioning (4 hits), vaginal application (2 hits), and oral food supplementation for glycemic control in diabetes mellitus type 2 (2 hits), and other conditions, including chronic kidney disease, stable angina pectoris, common cold, gastroenteritis, chronic thrombocytopenic purpura and mild cognitive impairment. Noteworthy, this search revealed only one study with published results. However, at present the majority of clinical trials are not listed in the publicly available specialized databases (Glanville et al. 2014). Concerning the published clinical trial data in research articles, most of the data on propolis efficacy in oral health applications is of poor methodological quality and doesn't allow drawing significant conclusions (Hwu and Lin 2014, Sung et al. 2017). The same conclusion is applicable for studies on skin and genital diseases (Sung et al. 2017). However, a meta-analysis of propolis treatment in cancer therapy-induced oral mucositis concludes that it is effective and safe (Kuo et al. 2018). Another meta-analysis concludes that the use of propolis as a supplement may be effective in the control of blood glucose concentrations in patients with type 2 diabetes mellitus (Karimian et al. 2019). The overview on clinical data points to the existence of sound interest on propolis pharmaceutical application, probably based on the tradition of apitherapy (Hellner et al. 2008). However, propolis products have been considered not sufficiently characterized in order to substantiate effects on health conditions (EFSA Panel on Dietetic Products and Allergies (NDA) 2010), which could also be among the reasons for the reported lack of reproducibility and poor methodological quality of clinical studies.

So far, the clinical potential of CAPE seems to remain relatively unexplored compared to propolis formulations. A quick search in the online database ClinicalTrials.gov with keywords caffeic acid phenethyl ester resulted in only one registered clinical study (ID:NCT02744703) on its properties as a matrix metalloproteinase inhibitor (Comlekoglu 2016). Not surprisingly, the pursuit towards finding applications in dentistry coincides with the abundance of clinical trials of total propolis extracts for dental application. The authors of the aforementioned study hypothesize that CAPE pretreatment of dentin would improve the binding of resin material to tooth substrate. The researchers summarize that dentin pretreatment with 5% CAPE solution "significantly increased the composite resin restorations' bond strengths to dentin applied with either total-etch or self-etch adhesive system". However,

Treatment	Condition/Procedure	Outcome	ClinicalTrials.gov Identifier, Phase and Status
CAPE solution (5%)	Restorative dentistry	Pretreatment improves bond strengths of nanohybrid resin restorations to dentin	NCT02744703; Completed
Single increasing oral doses of CC100 (CAPE in solvent 2 to 20 mg)	Healthy volunteers	No serious adverse events, $\mathrm{T_{max}}$ at 2.7h; $\mathrm{T_{1/2}}$ at 18.5 h	NCT02050334; Phase I Completed;
Multiple oral doses of CC100 (CAPE in solvent 250 to 1000 mg once daily)	Patients with amyotrophic lateral sclerosis	Anticipated results on safety, pharmacokinetics and pharmacodynamics	NCT03049046; Phase I; Recruiting;
Dexamethasone; Caffeic acid 300 mg, tablets, three times daily;	Patients with immune thrombocytopenia	Improves platelet count values; mild adverse effects	NCT02351622; Phase III; Completed; (Qin et al. 2015)
Dexamethasone; Caffeic acid 300 mg, tablets, three times daily;	Patients with immune thrombocytopenia	Anticipated results on sustained patient response after 6 months since the treatment started	NCT02556814; Phase IV; Unknown
Caffeic acid 300 mg, tablets, three times daily;	Patients with advanced esophageal squamous cell cancer	Anticipated results on 3 months' progression free survival and 1 year overall survival	NCT03070262; Phase III; Enrolling by invitation

Table 1. Clinical trials of CAPE and caffeic acid, listed in ClinicalTrials.org database as of 1.7.2019.

results' data is not yet publicly available. As caffeic acid is a metabolite of CAPE, we broadened the search in ClinicalTrials.gov database by applying the keywords "caffeic acid". This resulted in two more registered clinical trials of CAPE of synthetic origin, reconstituted in an unspecified diluent, administered orally. The first study, trialing the safety and tolerability of single increasing doses (2 to 20 mg), with dosing occurring every 2 to 7 days and a study duration of 5 to 15 days in healthy adults with a placebo group to which 2 increasing doses were administered and the third and highest dose was substituted with placebo (NCT02050334). The study has been completed and shows that there were no observed serious adverse events in none of the groups and there was no increase of the overall occurrence of other (not including-serious) adverse events in the high dose treatment group, compared to the lower dose treatment group, which confirms the notion of its tolerability from non-clinical data (Armutcu et al. 2015). The pharmacokinetic outcomes showed that the time to reach maximum observed plasma concentration (T_{max}) was on average 2.7 hours and the plasma decay halflife $(T_{1/2})$ was on average 18.5 hours. Probably, as a result of the favorable outcomes of this study, another clinical trial with the same CAPE formulation has been registered (NCT03049046) which has a status of recruiting subjects, suffering from amyotrophic lateral sclerosis for studying CAPE's multiple-dose safety and tolerability in doses up to 1000 mg. There have been numerous reports on the therapeutic potential of antioxidants as CAPE in the treatment of neurodegenerative diseases (Matteo and Esposito 2003, Fontanilla et al. 2012, Gandhi and Abramov 2012) This study plans to follow the effect of CAPE on some secondary outcome measures of pharmacodynamics, related to the condition of amyotrophic lateral sclerosis, namely the inflammation biomarker monocyte chemotactic protein 1 and the excitotoxicity/oxidative stress biomarkers heme oxygenase-1, thioredoxin and heat-shock protein 70. In an attempt to find non-listed in the searched database clinical trials, published in scientific articles, an additional literature search was undertaken in the following search engines: Scopus, Google Scholar and Crossref, applying as keywords "caffeic acid phenethyl ester" AND "clinical trial" by means of a software platform for bibliometric analysis (A. W. Harzing 2007). It failed to identify other clinical trials of CAPE.

The search in the ClinicalTrials.gov database with keywords "caffeic acid" resulted in two studies on the use of caffeic acid in the polypragmasy of immune thrombocytopenia (NCT02556814; NCT02351622) and one, applying caffeic acid for the treatment of esophageal cancer (NCT03070262). Unfortunately, no results are still available in the searched database. However, there is a publication, authored by the responsible party of both trials on the treatment of immune thrombocytopenia, reporting that treatment with 300 mg tablets of CAPE, three times daily for 12 weeks is "effective in patients with ITP with few and mild adverse effects" (Qin et al. 2015).

Conclusion

In conclusion, CAPE is the most researched and promising component of propolis, exerting most of the known pharmacological effects of total propolis extract. Due to its simple structure, it lacks the disadvantage of the inherently complex standardization of propolis. Historically, the initial phase in scientific research of CAPE, following its discovery, was marked by cornerstone research enterprises, providing evidence for its diverse biological activities. In the new millennium many of the mechanisms, underlying the biological effects of CAPE have been clarified, despite some important knowledge gaps. CAPE exerts its pharmacological activities by influencing the cellular signaling pathways, known to be in interplay with NF-κB and XPO1/CRM1. It is an effective inhibitor of enzymes, which are targeted by commercialized drugs and is a free radical scavenger. As information about the pharmacology of CAPE mounts, it is evident that it already attracting attention as a drug candidate molecule, which we exemplified with some recent clinical trials.

References

- Aggarwal BB, Sung B (2011) NF-κB in Cancer: A Matter of Life and Death. Cancer Discovery 1: 469–471. https://doi.org/10.1158/2159-8290.CD-11-0260
- Akyol S, Ozturk G, Ginis Z, Armutcu F, Yigitoglu MR, Akyol O (2013) In Vivo and In Vitro Antineoplastic Actions of Caffeic Acid Phenethyl Ester (CAPE): Therapeutic Perspectives. Nutrition and Cancer 65: 515–526. https://doi.org/10.1080/01635581.2013.776693
- Andrake MD, Skalka AM (2015) Retroviral integrase: then and now. Annual Review of Virology 2: 241–264. https://doi.org/10.1146/annurev-virology-100114-055043
- Aparov P, Kumar Reddy K, Reddanna P (2012) Structure and Ligand Based Drug Design Strategies in the Development of Novel 5-LOX Inhibitors. Current Medicinal Chemistry 19: 3763–3778. https://doi. org/10.2174/092986712801661112
- Arasoglu T, Derman S, Mansuroglu B (2016) Comparative evaluation of antibacterial activity of caffeic acid phenethyl ester and PLGA nanoparticle formulation by different methods. Nanotechnology 27: 025103. https://doi.org/10.1088/0957-4484/27/2/025103
- Armutcu F, Akyol S, Ustunsoy S, Turan FF (2015) Therapeutic potential of caffeic acid phenethyl ester and its anti-inflammatory and immunomodulatory effects (Review). Experimental and Therapeutic Medicine 9: 1582–1588. https://doi.org/10.3892/etm.2015.2346
- Bak J, Kim HJ, Kim SY, Choi Y-S (2016) Neuroprotective effect of caffeic acid phenethyl ester in 3-nitropropionic acid-induced striatal neurotoxicity. The Korean Journal of Physiology & Pharmacology: Official Journal of the Korean Physiological Society and the Korean Society of Pharmacology 20: 279–286. https://doi.org/10.4196/ kjpp.2016.20.3.279
- Baker RG, Hayden MS, Ghosh S (2011) NF-κB, inflammation and metabolic disease. Cell metabolism 13: 11–22. https://doi.org/10.1016/j. cmet.2010.12.008
- Bankova V, Trusheva B, Popova M (2018) Caffeic Acid Phenethyl Ester (CAPE) – Natural Sources, Analytical Procedures and Synthetic Approaches. "Prof. Marin Drinov" Publishing House of Bulgarian Academy of Sciences. https://doi.org/10.7546/CRABS.2018.09.01
- Baud V, Karin M (2009) Is NF-κB a good target for cancer therapy? Hopes and pitfalls. Nature Reviews. Drug Discovery 8: 33–40. https://doi.org/10.1038/nrd2781
- Berry CE, Hare JM (2004) Xanthine oxidoreductase and cardiovascular disease: molecular mechanisms and pathophysiological implications. The Journal of Physiology 555: 589–606. https://doi.org/10.1113/ jphysiol.2003.055913
- Carracedo A, Pandolfi PP (2008) The PTEN-PI3K pathway: of feedbacks and cross-talks. Oncogene 27: 5527–5541. https://doi.org/10.1038/ onc.2008.247
- Caton J, Ryan ME (2011) Clinical studies on the management of periodontal diseases utilizing subantimicrobial dose doxycycline (SDD). Pharmacological Research 63: 114–120. https://doi.org/10.1016/j.phrs.2010.12.003
- Celik S, Erdogan S, Tuzcu M (2009) Caffeic acid phenethyl ester (CAPE) exhibits significant potential as an antidiabetic and liver-protective agent in streptozotocin-induced diabetic rats. Pharmacological Research 60: 270–276. https://doi.org/10.1016/j.phrs.2009.03.017
- Center for Drug Evaluation and Research (2019) FDA grants accelerated approval to selinexor for multiple myeloma. FDA. http://www.fda. gov/drugs/resources-information-approved-drugs/fda-grants-accelerated-approval-selinexor-multiple-myeloma [July 8, 2019]

- Choi W, Villegas V, Istre H, Heppler B, Gonzalez N, Brusman N, Snider L, Hogle E, Tucker J, Oñate A, Oñate S, Ma L, Paula S (2019) Synthesis and characterization of CAPE derivatives as xanthine oxidase inhibitors with radical scavenging properties. Bioorganic Chemistry 86: 686–695. https://doi.org/10.1016/j.bioorg.2019.02.049
- Chung T-W, Moon S-K, Chang Y-C, Ko J-H, Lee Y-C, Cho G, Kim S-H, Kim J-G, Kim C-H (2004) Novel and therapeutic effect of caffeic acid and caffeic acid phenyl ester on hepatocarcinoma cells: complete regression of hepatoma growth and metastasis by dual mechanism. The FASEB Journal 18: 1670–1681. https://doi.org/10.1096/fj.04-2126com
- Comlekoglu MD (2016) Effect of Caffeic Acid Phenethyl Ester as a Matrix Metalloproteinase Inhibitor: Randomized Controlled Clinical Trial – No Study Results Posted – ClinicalTrials.gov. https://clinicaltrials.gov/ ct2/show/results/NCT02744703 [June 24, 2019]
- Dong X, Song Y-N, Liu W-G, Guo X-L (2009) MMP-9, a Potential Target for Cerebral Ischemic Treatment. Current Neuropharmacology 7: 269–275. https://doi.org/10.2174/157015909790031157
- Dow DE, Bartlett JA (2014) Dolutegravir, the Second-Generation of Integrase Strand Transfer Inhibitors (INSTIs) for the Treatment of HIV. Infectious Diseases and Therapy 3: 83–102. https://doi.org/10.1007/ s40121-014-0029-7
- Erdemli HK, Akyol S, Armutcu F, Akyol O (2015) Antiviral properties of caffeic acid phenethyl ester and its potential application. Journal of Intercultural Ethnopharmacology 4: 344–347. https://doi. org/10.5455/jice.20151012013034
- Fesen MR, Pommier Y, Leteurtre F, Hiroguchi S, Yung J, Kohn KW (1994) Inhibition of HIV-1 integrase by flavones, caffeic acid phenethyl ester (CAPE) and related compounds. Biochemical Pharmacology 48: 595–608. https://doi.org/10.1016/0006-2952(94)90291-7
- Flusberg DA, Sorger PK (2015) Surviving apoptosis: life-death signaling in single cells. Trends in cell biology 25: 446–458. https://doi. org/10.1016/j.tcb.2015.03.003
- Fontanilla CV, Wei X, Zhao L, Johnstone B, Pascuzzi RM, Farlow MR, Du Y (2012) Caffeic acid phenethyl ester extends survival of a mouse model of amyotrophic lateral sclerosis. Neuroscience 205: 185–193. https://doi.org/10.1016/j.neuroscience.2011.12.025
- Fuster JJ, Zuriaga MA, Ngo DT-M, Farb MG, Aprahamian T, Yamaguchi TP, Gokce N, Walsh K (2015) Noncanonical Wnt Signaling Promotes Obesity-Induced Adipose Tissue Inflammation and Metabolic Dysfunction Independent of Adipose Tissue Expansion. Diabetes 64: 1235–1248. https://doi.org/10.2337/db14-1164
- Gan K, Yang L, Xu L, Feng X, Zhang Q, Wang F, Tan W, Zhang M (2016) Iguratimod (T-614) suppresses RANKL-induced osteoclast differentiation and migration in RAW264.7 cells via NF-κB and MAPK pathways. International Immunopharmacology 35: 294–300. https:// doi.org/10.1016/j.intimp.2016.03.038
- Gandhi S, Abramov AY (2012) Mechanism of Oxidative Stress in Neurodegeneration. Oxidative Medicine and Cellular Longevity 2012: e428010. https://doi.org/10.1155/2012/428010
- Glanville JM, Duffy S, McCool R, Varley D (2014) Searching Clinical-Trials.gov and the International Clinical Trials Registry Platform to inform systematic reviews: what are the optimal search approaches? Journal of the Medical Library Association: JMLA 102: 177–183. https://doi.org/10.3163/1536-5050.102.3.007
- Grunberger D, Banerjee R, Eisinger K, Oltz EM, Efros L, Caldwell M, Estevez V, Nakanishi K (1988) Preferential cytotoxicity on tumor cells

by caffeic acid phenethyl ester isolated from propolis. Experientia 44: 230–232. https://doi.org/10.1007/BF01941717

- Günthard HF, Saag MS, Benson CA, del Rio C, Eron JJ, Gallant JE, Hoy JF, Mugavero MJ, Sax PE, Thompson MA, Gandhi RT, Landovitz RJ, Smith DM, Jacobsen DM, Volberding PA (2016) Antiretroviral Drugs for Treatment and Prevention of HIV Infection in Adults: 2016 Recommendations of the International Antiviral Society-USA Panel. JAMA 316: 191–210. https://doi.org/10.1001/jama.2016.8900
- Haines JD, Herbin O, de la Hera B, Vidaurre OG, Moy GA, Sun Q, Fung HYJ, Albrecht S, Alexandropoulos K, McCauley D, Chook YM, Kuhlmann T, Kidd GJ, Shacham S, Casaccia P (2015) Selective inhibitors of nuclear export avert progression in preclinical models of inflammatory demyelination. Nature Neuroscience 18: 511–520. https://doi.org/10.1038/nn.3953
- Hellner M, Winter D, Von Georgi R, Münstedt K (2008) Apitherapy: Usage and experience in German beekeepers. Evidence-based Complementary and Alternative Medicine 5: 475–479. https://doi. org/10.1093/ecam/nem052
- Hirsch GA, Bottomley PA, Gerstenblith G, Weiss RG (2012) Allopurinol acutely increases adenosine triphospate energy delivery in failing human hearts. Journal of the American College of Cardiology 59: 802–808. https://doi.org/10.1016/j.jacc.2011.10.895
- Hwu Y-J, Lin F-Y (2014) Effectiveness of propolis on oral health: a meta-analysis. The Journal of Nursing Research: JNR 22: 221–229. https://doi.org/10.1097/jnr.000000000000054
- Iochmann S, Bléchet C, Chabot V, Saulnier A, Amini A, Gaud G, Gruel Y, Reverdiau P (2009) Transient RNA silencing of tissue factor pathway inhibitor-2 modulates lung cancer cell invasion. Clinical & Experimental Metastasis 26: 457–467. https://doi.org/10.1007/s10585-009-9245-z
- Jin U-H, Chung T-W, Kang S-K, Suh S-J, Kim J-K, Chung K-H, Gu Y-H, Suzuki I, Kim C-H (2005) Caffeic acid phenyl ester in propolis is a strong inhibitor of matrix metalloproteinase-9 and invasion inhibitor: Isolation and identification. Clinica Chimica Acta 362: 57–64. https://doi.org/10.1016/j.cccn.2005.05.009
- Karimian J, Hadi A, Pourmasoumi M, Najafgholizadeh A, Ghavami A (2019) The efficacy of propolis on markers of glycemic control in adults with type 2 diabetes mellitus: A systematic review and meta-analysis. Phytotherapy Research: PTR 33: 1616–1626. https://doi. org/10.1002/ptr.6356
- Karin M (2009) NF-κB as a Critical Link Between Inflammation and Cancer. Cold Spring Harbor Perspectives in Biology 1. https://doi. org/10.1101/cshperspect.a000141
- Keum Y-S (2012) Regulation of Nrf2-Mediated Phase II Detoxification and Anti-oxidant Genes. Biomolecules & Therapeutics 20: 144–151. https://doi.org/10.4062/biomolther.2012.20.2.144
- Khoo KH, Verma CS, Lane DP (2014) Drugging the p53 pathway: understanding the route to clinical efficacy. Nature Reviews Drug Discovery 13: 217–236. https://doi.org/10.1038/nrd4236
- Kim H, Kim W, Yum S, Hong S, Oh J-E, Lee J-W, Kwak M-K, Park EJ, Na DH, Jung Y (2013) Caffeic acid phenethyl ester activation of Nrf2 pathway is enhanced under oxidative state: Structural analysis and potential as a pathologically targeted therapeutic agent in treatment of colonic inflammation. Free Radical Biology and Medicine 65: 552– 562. https://doi.org/10.1016/j.freeradbiomed.2013.07.015
- Kucukgul A (2016) Inhibition of Cigarette Smoke Induced-inflammation and Oxidative Damage by Caffeic Acid Phenethyl Ester in A549 Cells. Asian Journal of Pharmaceutics (AJP) 10. https://doi. org/10.22377/ajp.v10i04.913

- Kuo C-C, Wang R-H, Wang H-H, Li C-H (2018) Meta-analysis of randomized controlled trials of the efficacy of propolis mouthwash in cancer therapy-induced oral mucositis. Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer 26: 4001–4009. https://doi.org/10.1007/s00520-018-4344-5
- Lalor PF, Sun PJ, Weston CJ, Martin-Santos A, Wakelam MJO, Adams DH (2007) Activation of vascular adhesion protein-1 on liver endothelium results in an NF-κB-dependent increase in lymphocyte adhesion. Hepatology 45: 465–474. https://doi.org/10.1002/hep.21497
- Lee Y, Shin D, Kim J-H, Hong S, Choi D, Kim Y-J, Kwak M-K, Jung Y (2010) Caffeic acid phenethyl ester-mediated Nrf2 activation and IκB kinase inhibition are involved in NFκB inhibitory effect: Structural analysis for NFκB inhibition. European Journal of Pharmacology 643: 21–28. https://doi.org/10.1016/j.ejphar.2010.06.016
- Lee Y-J, Kuo H-C, Chu C-Y, Wang C-J, Lin W-C, Tseng T-H (2003) Involvement of tumor suppressor protein p53 and p38 MAPK in caffeic acid phenethyl ester-induced apoptosis of C6 glioma cells. Biochemical Pharmacology 66: 2281–2289. https://doi.org/10.1016/j. bcp.2003.07.014
- Li H, Qiu Z, Li F, Wang C (2017) The relationship between MMP-2 and MMP-9 expression levels with breast cancer incidence and prognosis. Oncology Letters 14: 5865–5870. https://doi.org/10.3892/ ol.2017.6924
- Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, Gargiulo G, Testa G, Cacciatore F, Bonaduce D, Abete P (2018) Oxidative stress, aging, and diseases. Clinical Interventions in Aging 13: 757–772. https://doi.org/10.2147/CIA.S158513
- Lingappan K (2018) NF-κB in Oxidative Stress. Current Opinion in Toxicology 7: 81–86. https://doi.org/10.1016/j.cotox.2017.11.002
- Liu G-L, Han N-Z, Liu S-S (2018) Caffeic acid phenethyl ester inhibits the progression of ovarian cancer by regulating NF-?B signaling. Biomedicine & Pharmacotherapy = Biomedecine & Pharmacotherapie 99: 825–831. https://doi.org/10.1016/j.biopha.2018.01.129
- Loboda A, Damulewicz M, Pyza E, Jozkowicz A, Dulak J (2016) Role of Nrf2/HO-1 system in development, oxidative stress response and diseases: an evolutionarily conserved mechanism. Cellular and Molecular Life Sciences 73: 3221–3247. https://doi.org/10.1007/s00018-016-2223-0
- Ma B, Hottiger MO (2016) Crosstalk between Wnt/β-Catenin and NFκB Signaling Pathway during Inflammation. Frontiers in Immunology 7. https://doi.org/10.3389/fimmu.2016.00378
- Machlus KR, Wu SK, Vijey P, Soussou TS, Liu Z-J, Shacham E, Unger TJ, Kashyap T, Klebanov B, Sola-Visner M, Crochiere M, Italiano JE, Landesman Y (2017) Selinexor-induced thrombocytopenia results from inhibition of thrombopoietin signaling in early megakaryopoiesis. Blood 130: 1132–1143. https://doi.org/10.1182/ blood-2016-11-752840
- Martins R, Lithgow GJ, Link W (2016) Long live FOXO: unraveling the role of FOXO proteins in aging and longevity. Aging Cell 15: 196– 207. https://doi.org/10.1111/acel.12427
- Matteo V, Esposito E (2003) Biochemical and Therapeutic Effects of Antioxidants in the Treatment of Alzheimers Disease, Parkinsons Disease, and Amyotrophic Lateral Sclerosis. Current Drug Target – CNS & Neurological Disorders 2: 95–107. https://doi. org/10.2174/1568007033482959
- Michaluart P, Masferrer JL, Carothers AM, Subbaramaiah K, Zweifel BS, Koboldt C, Mestre JR, Grunberger D, Sacks PG, Tanabe T (1999) Inhibitory effects of caffeic acid phenethyl ester on the activity and

expression of cyclooxygenase-2 in human oral epithelial cells and in a rat model of inflammation. Cancer Research 59: 2347–2352.

- Moynagh PN (2005) The NF-κB pathway. Journal of Cell Science 118: 4589–4592. https://doi.org/10.1242/jcs.02579
- Murtaza G, Karim S, Akram MR, Khan SA, Azhar S, Mumtaz A, Bin Asad MHH (2014) Caffeic Acid Phenethyl Ester and Therapeutic Potentials. BioMed Research International 2014. https://doi. org/10.1155/2014/145342
- Natarajan K, Singh S, Burke TR, Grunberger D, Aggarwal BB (1996) Caffeic acid phenethyl ester is a potent and specific inhibitor of activation of nuclear transcription factor NF-kappa B. Proceedings of the National Academy of Sciences 93: 9090–9095. https://doi. org/10.1073/pnas.93.17.9090
- Qin P, Wei Y, Hou M, Zhao C, Shen Z (2015) A multicenter clinical trial of caffeic acid tablet in treatment of 103 primary immune thrombocytopenia patients. Zhonghua Xue Ye Xue Za Zhi = Zhonghua Xueyexue Zazhi 36: 103–106.
- Razzaghi-Asl N, Garrido J, Khazraei H, Borges F, Firuzi O (2013) Antioxidant Properties of Hydroxycinnamic Acids: A Review of Structure- Activity Relationships. Current Medicinal Chemistry 20: 4436– 4450. https://doi.org/10.2174/09298673113209990141
- Rossi A, Longo R, Russo A, Borrelli F, Sautebin L (2002) The role of the phenethyl ester of caffeic acid (CAPE) in the inhibition of rat lung cyclooxygenase activity by propolis. Fitoterapia 73: S30–S37. https:// doi.org/10.1016/S0367-326X(02)00188-0
- Salminen A, Ojala J, Huuskonen J, Kauppinen A, Suuronen T, Kaarniranta K (2008) Interaction of aging-associated signaling cascades: Inhibition of NF-κB signaling by longevity factors FoxOs and SIRT1. Cellular and Molecular Life Sciences 65: 1049–1058. https://doi. org/10.1007/s00018-008-7461-3
- Schneider G, Krämer OH (2011) NFκB/p53 crosstalk a promising new therapeutic target. Biochimica et Biophysica Acta (BBA) – Reviews on Cancer 1815: 90–103. https://doi.org/10.1016/j.bbcan.2010.10.003
- Shigeoka Y, Igishi T, Matsumoto S, Nakanishi H, Kodani M, Yasuda K, Hitsuda Y, Shimizu E (2004) Sulindac sulfide and caffeic acid phenethyl ester suppress the motility of lung adenocarcinoma cells promoted by transforming growth factor-beta through Akt inhibition. Journal of Cancer Research and Clinical Oncology 130: 146– 152. https://doi.org/10.1007/s00432-003-0520-0
- Skiles JW, Gonnella NC, Jeng AY (2004) The design, structure, and clinical update of small molecular weight matrix metalloproteinase inhibitors. Current Medicinal Chemistry 11: 2911–2977. https://doi. org/10.2174/0929867043364018
- Stähli A, Maheen CU, Strauss FJ, Eick S, Sculean A, Gruber R (2019) Caffeic acid phenethyl ester protects against oxidative stress and dampens inflammation via heme oxygenase 1. International Journal of Oral Science 11: 6. https://doi.org/10.1038/s41368-018-0039-5
- Sud'ina GF, Mirzoeva OK, Pushkareva MA, Korshunova GA, Sumbatyan NV, Varfolomeev SD (1993) Caffeic acid phenethyl ester as a lipoxygenase inhibitor with antioxidant properties. FEBS Letters 329: 21– 24. https://doi.org/10.1016/0014-5793(93)80184-V
- Sung S-H, Choi G-H, Lee N-W, Shin B-C (2017) External Use of Propolis for Oral, Skin, and Genital Diseases: A Systematic Review and Meta-Analysis. Evidence-Based Complementary and Alternative Medicine. https://doi.org/10.1155/2017/8025752

- Tolba MF, Omar HA, Azab SS, Khalifa AE, Abdel-Naim AB, Abdel-Rahman SZ (2016) Caffeic Acid Phenethyl Ester: A Review of Its Antioxidant Activity, Protective Effects against Ischemia-reperfusion Injury and Drug Adverse Reactions. Critical Reviews in Food Science and Nutrition 56: 2183–2190. https://doi.org/10.1080/10408398.2013.821967
- Tolba MF, Esmat A, Al-Abd AM, Azab SS, Khalifa AE, Mosli HA, Abdel-Rahman SZ, Abdel-Naim AB (2013) Caffeic acid phenethyl ester synergistically enhances docetaxel and paclitaxel cytotoxicity in prostate cancer cells. IUBMB Life 65: 716–729. https://doi. org/10.1002/iub.1188
- Tseng J-C, Lin C-Y, Su L-C, Fu H-H, Yang S-D, Chuu C-P (2016) CAPE suppresses migration and invasion of prostate cancer cells via activation of non-canonical Wnt signaling. Oncotarget 7: 38010–38024. https://doi.org/10.18632/oncotarget.9380
- Turan I, Demir S, Misir S, Kilinc K, Mentese A, Aliyazicioglu Y, Deger O (2015) Cytotoxic Effect of Turkish Propolis on Liver, Colon, Breast, Cervix and Prostate Cancer Cell Lines. Tropical Journal of Pharmaceutical Research 14: 777. https://doi.org/10.4314/tjpr.v14i5.5
- Urushibara M, Takayanagi H, Koga T, Kim S, Isobe M, Morishita Y, Nakagawa T, Löeffler M, Kodama T, Kurosawa H, Taniguchi T (2004) The antirheumatic drug leflunomide inhibits osteoclastogenesis by interfering with receptor activator of NF-kB ligand–stimulated induction of nuclear factor of activated T cells c1. Arthritis & Rheumatism 50: 794–804. https://doi.org/10.1002/art.20206
- Wang AY, Liu H (2019) The past, present, and future of CRM1/XPO1 inhibitors. Stem Cell Investigation 6. https://doi.org/10.21037/ sci.2019.02.03
- Wang L-C, Chu K-H, Liang Y-C, Lin Y-L, Chiang B-L (2010) Caffeic acid phenethyl ester inhibits nuclear factor-κB and protein kinase B signalling pathways and induces caspase-3 expression in primary human CD4+ T cells. Clinical and Experimental Immunology 160: 223–232. https://doi.org/10.1111/j.1365-2249.2009.04067.x
- Wang S-H, Chen C-S, Huang S-H, Yu S-H, Lai Z-Y, Huang S-T, Lin C-M (2009) Hydrophilic ester-bearing chlorogenic acid binds to a novel domain to inhibit xanthine oxidase. Planta Medica 75: 1237–1240. https://doi.org/10.1055/s-0029-1185521
- Wardyn JD, Ponsford AH, Sanderson CM (2015) Dissecting molecular cross-talk between Nrf2 and NF-κB response pathways. Biochemical Society Transactions 43: 621–626. https://doi.org/10.1042/ BST20150014
- Watabe M, Hishikawa K, Takayanagi A, Shimizu N, Nakaki T (2004) Caffeic Acid Phenethyl Ester Induces Apoptosis by Inhibition of NFκB and Activation of Fas in Human Breast Cancer MCF-7 Cells. Journal of Biological Chemistry 279: 6017–6026. https://doi.org/10.1074/jbc.M306040200
- Zabaiou N, Fouache A, Trousson A, Baron S, Zellagui A, Lahouel M, Lobaccaro J-MA (2017) Biological properties of propolis extracts: Something new from an ancient product. Chemistry and Physics of Lipids 207: 214–222. https://doi.org/10.1016/j.chemphyslip.2017.04.005
- Zhan T, Rindtorff N, Boutros M (2017) Wnt signaling in cancer. Oncogene 36: 1461–1473. https://doi.org/10.1038/onc.2016.304
- Zhang P, Tang Y, Li N-G, Zhu Y, Duan J-A (2014) Bioactivity and Chemical Synthesis of Caffeic Acid Phenethyl Ester and Its Derivatives. Molecules 19: 16458–16476. https://doi.org/10.3390/molecules191016458